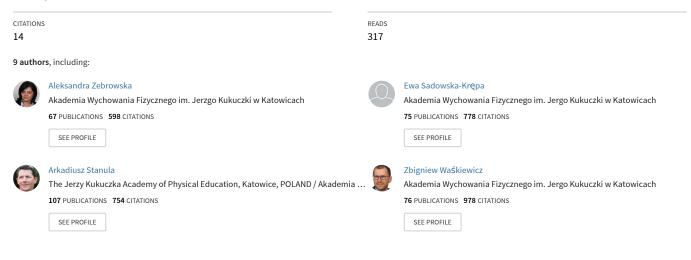
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## The Effect of Vitamin D Supplementation on Serum Total 25(OH)D Levels and Biochemical Markers of Skeletal Muscles in Runners Vitamin D Supplementation in Marathon Runners

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## Journal of the International Society of Sports Nutrition

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Question	Response
<b>Is this study a clinical trial?</b> <hr/> <i>A clinical trial is defined by the World Health Organisation as 'any research study that prospectively assigns human participants or groups of humans to one or more health-related interventions to evaluate the effects on health outcomes'.</i>	No

1	The Effect of Vitamin D Supplementation on Serum Total 25(OH)D
2	Levels and Biochemical Markers of Skeletal Muscles in Runners
3	
4	Vitamin D Supplementation in Marathon Runners
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#### 33 Abstract

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The study aimed to evaluate the effects of a 3-week vitamin D supplementation on serum 35 25(OH)D levels and skeletal muscle biomarkers (*i.e.* troponin, myoglobin, creatine kinase and lactic dehydrogenase) of endurance runners. Twenty-four runners were examined at 36 37 baseline and in response to eccentric exercise before and after two dietary protocols (dose 38 of 2000 IU for three weeks or placebo). Significant differences between pre- and post-39 intervention in 25(OH)D levels were observed (36.1 $\pm$ 6.0 versus 40.0 $\pm$ 5.2 ng/ml, p<0.05). 40 A higher post intervention 25(OH)D level was observed after vitamin D diet compared to 41 placebo (40.0±5.2 versus 31.8±4.2 ng/mL, respectively; p<0.01). The vitamin D 42 supplementation decreased 1 h and 24 h post-exercise troponin (p < 0.05, p < 0.01, 43 respectively), myoglobin concentration (p < 0.05, p < 0.01, respectively) and 24 h post 44 exercise creatine kinase (CK) activity (p < 0.01). A negative correlation was observed 45 between post exercise 25(OH)D levels and myoglobin levels (r=-0.57; p<0.05), 25(OH)D 46 levels and CK (r=-0.60; p<0.05), and 25(OH)D levels and TNF $\alpha$  (r=-0.58; p<0.05). These 47 findings suggested that an increase in 25(OH)D release in response to vitamin D 48 supplementation attenuated the muscle biomarker levels following eccentric exercise and 49 might play a key role in prevention of skeletal muscle injury.

50 Key words: vitamin D; muscle biomarkers; eccentric exercise; fatigue; marathon.

#### 51 Introduction

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53 Strenuous exercise has been associated with adaptive changes in skeletal muscle, such as an 54 ability to use oxygen to generate energy for muscle work, a decrease in oxygen demand for 55 the same level of external work performed, as well as an improvement of mechanisms towards decreased exercise-induced muscle damage<sup>1</sup>. In a recent study, a prevalence of 56 57 vitamin D deficiency in extreme endurance athletes, and an association between delayed 58 physical performance and the deficiency in vitamin D were observed during regular training <sup>2-4</sup>. These physiological responses in muscles were influenced by exercise-induced 59 60 mechanisms and were probably affected by nutritional athletic status and limitation of sun exposure <sup>2, 5-7</sup>. 61

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Long distance running has been shown to induce progressive increase of neuromuscular 63 64 function and adaptive changes in cardiovascular, as well as immune and endocrine systems  $^{8-11}$ . The potential mechanisms - through which function of the muscular system might be 65 66 beneficially modified in response to extreme repeated exercise stress - included improvement of vitamin D status<sup>4</sup>. Several studies supported the theory that functional 67 68 responses in skeletal muscle were influenced by mechanisms that could be affected by 69 biological effects of an active form of vitamin D and its ability to bind with the membrane and nuclear vitamin D receptors (VDRs)<sup>11, 12</sup>. Besides the importance of vitamin D, 70 71 especially 25(OH)D (serum 25-hydroxy vitamin D), in the regulation of bones and calcium 72 homeostasis, it was also involved in skeletal muscle performance and in exercise-induced inflammatory processes, neurological functions and cardiovascular health <sup>7, 13-15</sup>. It should 73 74 be noted that muscle power and force in marathon runners were linked with vitamin D levels <sup>16</sup>. The deficiency in vitamin D increased the risk of muscle myopathy, and impaired 75 cross-bridge formation leading to muscle weakness and fatigue <sup>17-19</sup>. Due to the higher 76

77 levels of biomarkers of muscle injury and reduction of total antioxidant capacity and 78 muscle function in response to extreme exercise training, strategies should be developed to 79 maintain an optimal vitamin D level in response to its exercise-induced deficiency. It has 80 been hypothesized that higher exposure to vitamin D - producing ultraviolet light and 81 serum 25(OH)D levels above the normal reference range (up to 50 ng/mL) - could be 82 associated with beneficial adaptations in skeletal muscle consisting of enhanced aerobic 83 performance, both force and power production and decreased recovery time from training 20 84

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86 The physiological consequence of intense physical training in response to vitamin D 87 supplementation induced by activation of the serum 25(OH)D status depended on the dosages exceeding the recommendations for vitamin D<sup>21-24</sup>. In elite rowers, maximal 88 89 oxygen uptake increased significantly in response to supplementation with 6000 IU/day of 90 vitamin D during 8-weeks training, whereas, the dosage of 4000 IU/day for 35 days of 91 vitamin D improved the recovery by the attenuation of the inflammation processes in 92 moderately active adults <sup>25</sup>. Positive effects of supplementation (8 weeks of 5000 IU/day of 93 vitamin D) and increases in force and power production in professional soccer players were also observed <sup>24</sup>. However, optimal vitamin D dosage and serum levels needed for athletic 94 95 performance and recovery have been controversial <sup>25</sup>. A dosage of 600-800 IU/day and 96 1000 IU/day of vitamin D might not be sufficient for optimal levels of vitamin D, nor prevent a decline in serum 25(OH)D in response to intense exercise training <sup>21</sup>. There was 97 98 evidence suggesting that dietary supplementation with 2000 to 5000 IU/day of vitamin D 99 had a positive impact on bone health and skeletal muscle function <sup>23</sup>. However, it was not 100 specified what dose of vitamin D was sufficient to prevent muscle damage and could be 101 effective for accelerating muscle regeneration after intense effort with an eccentric work component <sup>26, 27</sup>. 102

Participation in marathon and ultra-marathon races is becoming an increasingly popular
activity, which is encouraged by an increasing number of running events being organized
each year. Hence, a number of investigations have been conducted to determine the risk
factors of skeletal muscle injury in long-term runners <sup>9, 28</sup>. Considering that fact, there are
still, at present, no official recommendations for the treatment of muscle fatigue.
Nonspecific treatments with higher vitamin D usage have been used clinically or
experimentally, and have shown some positive effects.

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111 Therefore, it seemed important to investigate the association between recommended low 112 vitamin D dosage and an early identification of increased muscle fatigue risk. In previous 113 studies on the assessment of muscle dysfunction, the conventional biomarkers (e.g., Tn, CK, myoglobin, LDH) have been analyzed <sup>29, 30</sup>. These markers had different release times 114 and different times of reaching maximal concentrations <sup>8, 10, 31</sup>. It has been hypothesized that 115 116 exercise-induced lower muscle biomarker secretion may depend on increased serum 117 25(OH)D levels and these vitamin levels might be used for early detection of greater 118 muscle resistance to fatigue. There are limited data regarding the effect of lower dosages of 119 vitamin D supplementation on muscle function and optimization of recovery mechanisms 120 of elite ultramarathon runners. It was also hypothesized that higher serum 25(OH)D levels in response to low dosage of vitamin D supplementation might improve this function via 121 122 the stimulation of 25(OH)D production and release. To verify this, the relationships 123 between eccentric exercise-induced muscle biomarker levels, as measured by troponin, 124 myoglobin concentrations and creatine kinase and lactic dehydrogenase activity and 125 25(OH)D levels in response to vitamin D supplementation in marathon runners were 126 examined.

#### 128 Material and Methods

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#### 130 **Ethical approval**

- 131 The experiment was approved by the Ethics Committee of the Academy of Physical
- 132 Education in Katowice (Ethics Committee decision KBN 3.2016) and conformed to the

133 standards set by the Declaration of Helsinki.

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#### 135 Subjects

136 Twenty-four male ultramarathon runners who were endurance-trained for about seven years 137 participated in the study. They were randomly assigned to either dietary protocol (*i.e.* 138 placebo or the vitamin D supplementation, placebo-controlled study). All subjects 139 participated in the study during the pre-season period. Study members were recruited from 140 all the competitors of the ultra-marathons held during the Polish Running Championships. 141 The inclusion criteria were participation in at least five marathons and written informed 142 consent to take part in the study. The training status of the subjects included in the 143 supplemented and placebo group expressed as maximal oxygen consumption (VO<sub>2</sub>max) 144 was 54.5±9.4 and 50.1±7.4 ml/kg/min, respectively. Age, height, body mass, body mass 145 index (BMI) and body composition of the participants (Mean±SD) are presented in Table 1. 146 Mean energy supply with diet, mean daily fat, carbohydrate, protein and vitamin D intake 147 were comparable in the supplemented group and placebo group (Table 2). Biochemical 148 measurements of pre intervention 25(OH)D levels in runners indicate that serum levels of 149 25(OH)D did not differ between the groups (Table 3). 150

151 All subjects reported that they were not taking any medication that could affect the

152 25(OH)D status. They were instructed to abstain from strenuous exercise for 24 hours

153 before the ultrasound measurements. No caffeine, supplements, or alcohol were permitted 154 during the 48 hours before the experiment. Three weeks prior to the study all participants 155 were put on a mixed diet (Table 2). The composition of the diet was calculated with 156 dedicated software for each subject (Dietus, B.U.I. InFit. Warsaw, Poland). The diet was 157 continued with vitamin D or placebo administration. To ensure that participants adhered to 158 the dietary regimen, they had to keep daily food intake logs which were inspected during 159 the weekly, obligatory visits in the laboratory. We supplemented our subjects for 3 weeks 160 and before each diet protocol, the biochemical variables and physiological variables were 161 analyzed.

#### 162 Supplementation procedure and training protocol

All clinical data, including biochemical parameters and exercise examination, were obtained after an overnight fast. Following these measurements, blood samples were taken through a peripheral catheter inserted into the antecubital vein; each participant completed an incremental ergometer exercise test. After initial testing, the vitamin D supplemented group received 50  $\mu$ g (2 x 1000 IU/day) of vitamin D. The control group received a placebo in the form of gelatin capsules (1.3 g lactose monohydrate). Participants were instructed to take the capsules with meals twice daily for a total of 3 weeks.

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#### 171 Exercise protocols

All subjects participated in the following experiment consisting of three protocols: the
incremental exercise test (to determine the intensity of continuous eccentric exercise,
downhill running) continuous eccentric exercise before supplementation and continuous
eccentric exercise post supplementation (preExE and postExE, respectively). The two
laboratory protocols were separated by at least seven days to prevent any possible

177 interference on the subjects' exercise abilities or fatigue. At the baseline, before treatment 178 protocol (supplementation or placebo), all subjects performed a standard incremental 179 treadmill exercise test (LE 200 treadmill, Jaeger, Frankfurt, Germany) to measure their 180 individual aerobic performance (maximal oxygen uptake, VO2max). The test started with a 181 3-min warm-up at 6 km/h and 0° inclination; the intensity was then increased by 2 km/h 182 every 3 min up to 12 km/h and then the intensity was increased and inclination by 2.5° up 183 to maximal exercise intensity or volitional fatigue. Heart rate (HR) (PE-3000 Sport-Tester, 184 Polar Inc., Kempele, Finland) and systolic and diastolic blood pressure (SBP/DBP) were 185 measured (HEM-907 XL, Omron Corporation, Kyoto, Japan) before and immediately after 186 the test. Pulmonary ventilation (VE), oxygen uptake (VO<sub>2</sub>), and carbon dioxide output 187 (CO<sub>2</sub>) were measured continuously from the 6 minutes prior to exercise test and throughout 188 each stage of the exercise test using the Oxycon Apparatus (CareFusion Germany 234 189 GMBH, Hoechberg Jaeger, Germany). Physiological characteristics of the participants are 190 presented in Table 1.

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192 In the second phase of the study, the subjects participated in a 30-minuterunning test with 193 an eccentric type of work (ExE) and intensity of their individual 70% VO<sub>2</sub>max and 194 treadmill 16° inclination based on a modified test protocol (AR Young Company, 195 Indianapolis)<sup>32</sup>. According to Sorichter *et al.* <sup>32</sup>, it has been shown that running down, i.e. 196 eccentric effort, is an effective way to cause such a load on skeletal muscle that it can 197 induce delayed onset muscle soreness (DOMS) symptoms. All subjects participated in the 198 third laboratory protocol after 3 weeks of vitamin D supplementation or placebo according 199 to the same ExE protocol.

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#### 202 Measurements and blood collection

At the beginning of the study (pre intervention) and at the end of each treatment period 203 204 (post intervention supplementation or placebo protocol) all subjects reported to the 205 laboratory and had venous blood drawn for the determination of levels of 25(OH)D and 206 muscle biomarker concentrations. The blood samples were collected to determine the 207 aforementioned markers immediately before (rest), immediately after the eccentric exercise 208 (max) and during post-workout recovery (60 min and 24 hours after the end of the test).All 209 investigated subjects underwent bioelectric impedance analysis (InBody Data Management 210 System) under resting conditions to determine their body mass. The exercise tolerance was 211 assessed by heart rate (HR) and blood lactate concentrations (LA) in response to eccentric 212 exercise.

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#### 214 **Biochemical analyses**

215 For biochemical analysis, antecubital venous blood samples were always drawn at the same 216 time of day, with the subject in a seated position. Venous blood samples were collected at 217 four time points. Blood was allowed to clot at room temperature and then centrifuged. The 218 resulting serum was aliquoted and frozen at -80°Cfor later analyses. The measurements of 219 serum 25(OH)D levels were performed using25OH- Vitamin D ImmunoAssay(DIA source 220 25OH Vitamin D total RIA CT Kit, Belgium). Intra- and interassay coefficients 221 of variation for 25(OH)D were 5.9 - 3.3 % and 7.4 - 4.9 %, respectively. The measurements 222 of troponin (TN) were performed using Human TNNI1 (Troponin I Type 1, Slow Skeletal 223 ELISA Kit EH-0625, Fine Biological Technology, Co Ltd. Wuhan, China). Intra- and 224 interassay coefficients of variation for TN were < 8.0 % and < 10.0 %, respectively. The 225 serum myoglobin (MB) levels were measured using Human Myoglobin Enzyme 226 Immunoassay (Mioglobina ELISA, KIT DRG® Myoglobin, EIA-3955). Intra- and 227 interassay coefficients of variation for MB were 3.9 - 6.6% and 7.8 - 7.2%, respectively.

228 The lowest detectable level of myoglobin by this assay is estimated to be 5 ng/ml. The 229 proinflammatory cytokines interleukin-6 (IL-6) levels were measured by using Human IL-6 230 High Sensitive ELISA kit, Diacone, France. Intra- and inter-assay coefficients of variation 231 for of IL-6 were < 4.4% and < 6.4%, respectively and tumor necrosis factor-alpha (TNF- $\alpha$ ) 232 were performed using (TNF-α-EASIA KAP1751 firm DIAsource, Belgium). Intra- and 233 interassay coefficients of variation for TNF- $\alpha$  were < 5.1 % and < 8.6 %, %, respectively. 234 Creatine Kinase (CK) and Lactate Dehydrogenase (LDH) activity were measured using a 235 commercial kit (CK NAC and LDH P-L, RANDOX, UK). Intra- and interassay coefficients 236 of variation for CK were 2.3 - 1.5 % and 3.9 - 3.3%, respectively and for LDH were 3.9 -237 1.8 % and 4.0 - 2.8%, respectively. Blood lactate concentrations (LA) were determined 238 using BiosenC\_line method (EKF Diagnostic GmbH, Germany). The degree of 239 hemoconcentration (%) was calculated according to formula of subtracting the peak 240 hematocrit with the minimum hematocrit recorded and multiplying by 100; all biochemical

241 variables levels were corrected according to plasma volume.

#### 242 Statistical Analysis

243 Shapiro-Wilk, Levene's and Mauchly's tests were used in order to verify the normality, 244 homogeneity and sphericity of the sample's data variances, respectively. The magnitudes of 245 differences between results of pre-test and post-test were expressed as a standardized mean 246 difference (Cohen effect sizes). The criteria to interpret the magnitude of the effect sizes 247 were: <0.2 trivial, 0.2—0.6 small, 0.6—1.2 moderate, 1.2—2.0 large and >2.0 very large. 248 Descriptive statistics were calculated and the results were presented as means and standard 249 deviations (mean±SD). We analyzed differences between pre- and post-intervention 250 (placebo/vitamin D) baseline and post exercise variables. The data were analyzed by two-251 way ANOVA followed by the Student-Newman-Keuls test when appropriate. The 252 statistical analysis includes a two-way ANOVA (placebo vs. vitamin D) and pre 253 intervention vs. post intervention. Pearson correlation coefficients were analyzed to

254	determine the inter-variable relationships. All analyses were performed using the Statistica
255	v. 12 statistical software package (StatSoft, Tulsa, OK, USA). Statistical significance was
256	set at <i>p</i> <0.05.
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277 **Results** 

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279 The effects of dietary supplementation with vitamin D and placebo administration on serum 280 25(OH)D, muscle biomarkers and proinflammatory cytokines concentrations in runners 281 were compared after three weeks of each treatment protocol. Analysis of variance revealed 282 a significant effect of vitamin D supplementation on serum 25(OH)D concentration 283 (F=17.1; p<0.001). Significant differences between pre-intervention and post-intervention 284 baseline serum 25(OH)D levels (p<0.05) and post ExE levels were observed after the 285 vitamin D dietary protocol (p<0.001). A significantly higher post intervention baseline 286 25(OH)D level was observed after vitamin D diet compared to placebo (40.0±5.2 versus 287 31.8±4.2 ng/ml, p < 0.05, respectively). The vitamin D increased baseline 25(OH)D ( $\Delta$ ) by 288 5.7±2.8 ng/ml and decreased placebo by - 2.2±3.6 ng/ml. ANOVA revealed a significant 289 effect of vitamin D diet on TN levels (F=11.6; p<0.01). A significantly lower 24 h post 290 exercise TN level was observed in vitamin D diet compared to pre-supplementation values 291 (p < 0.05). The baseline and max TN levels were significantly lower in vitamin D diet 292 compared to placebo (p < 0.05 and p < 0.001, respectively). A significant effect of vitamin D 293 supplementation was observed in response to MB levels (F=9.0; p<0.01) and TNF $\alpha$  (F=4.7; 294 p < 0.05). A repeated measure of two-way ANOVA revealed the significance of diet and 295 exercise interaction effects on MB (F=4.5; p<0.01), CK (F=4.5; p<0.01) and 25(OH)D 296 concentration (F=3.2; p < 0.05). 297

A significantly lower 24h post ExE CK activity was observed after vitamin D diet compared to the pre intervention and placebo group (p<0.05 and p<0.05, respectively). No significant effect of vitamin D diet was observed regarding LDH activity at baseline and at post-exercise levels. Significant lower max and 1h post ExE TNF $\alpha$  levels were observed

302	after vitamin D diet compared to pre-intervention ( $p$ <0.01 and $p$ <0.01, respectively) and a
303	non-significant trend to lower IL-6 levels (Table 3).

305	A significant and negative correlation was observed between 25(OH)D concentration and
306	TN level (24 h post ExE) in response to supplementation (r=-0.49; $p$ <0.05) and 25(OH)D
307	(Figure 1) and MB concentration (r=- $0.57$ ; $p=0.05$ ) (Figure 2). Importantly, the negative
308	correlation was observed between 25(OH)D concentration and CK activity during the 24h
309	recovery period (r=-0.60; $p$ <0.05) and TNF $\alpha$ levels (r=-0.42; $p$ <0.05) (Figure 3) only in
310	response to vitamin D supplementation. ANOVA did not reveal any significant effect of
311	diet on HRmax (157.0±5.0 <i>versus</i> 154.0±3.0 b/min) and serum LA (1.9±0.3 <i>versus</i> 1.8±0.3)
312	concentrations in response to ExE ( $p$ >0.05).
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325 **Discussion** 

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327 The present study was undertaken to investigate whether vitamin D supplementation might 328 exert a beneficial effect on serum 25(OH)D concentrations, skeletal muscle biomarkers, and 329 an exercise tolerance in marathon runners. Our results have demonstrated that a three-week 330 low dosage of vitamin D supplementation caused elevation of baseline serum 25(OH)D 331 compared to pre-supplementation levels. An increase in baseline and post-exercise serum 332 25(OH)D were also observed in contrast to the placebo administration. Moreover, the 333 increased 25(OH)D production seem to have significant effect on resting and post eccentric 334 exercise - induced skeletal biomarker levels and proinflammatory cytokines. The major 335 findings of our study are that greater 25(OH)D expression in response to vitamin D diet 336 correlated with biomarkers of muscle damage and that this effect is more pronounced 337 during 24h recovery. Three weeks of supplementation had a beneficial effect on skeletal 338 muscle function. Lower serum levels of biomarkers of skeletal muscle damage and vitamin 339 D status improvement might, in turn, have significantly decreased individual recovery time 340 from eccentric exercise.

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342 Data concerning positive impacts of vitamin D consumption on optimizing athletic performance and recovery in intensely trained athletes are still sparse <sup>5, 20, 33</sup>. Most studies 343 344 support the benefits of dietary supplementation with vitamin D in healthy untrained adults and people diagnosed with 25-hydroxyvitamin D insufficiency (<30 ng/ml)<sup>24, 34, 35</sup>. These 345 results revealed a positive effect of vitamin D supplementation on global muscle strength. 346 power and mass <sup>14, 17, 36</sup>. Supplementation also seems more effective on people aged 65 347 348 vears compared to younger subjects. The effectiveness of the vitamin D supplementation 349 was confirmed in athletes, however, the optimal intake and serum 25(OH)D levels have yet to be identified in the athletic population<sup>2</sup>. In the study of Zhang *et al.*, vitamin D 350

supplementation positively affected lower limb muscle strength, but not muscle power in
athletes <sup>37</sup>. It has been suggested that different muscle groups may respond differently to
vitamin D supplementation. Significant improvements in muscle function following
vitamin D repletion were reported in a study on females<sup>38</sup>. Contrarily, a recent metaanalysis involving 532 athletes found no improvement in measures of physical performance
despite the inclusion of vitamin D deficient athletes at baseline and improvements in
vitamin D levels over mean 12 weeks of follow-up <sup>5</sup>.

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It has recently been reported that vitamin D supplementation might influence aerobic performance in athletes <sup>36, 39</sup>. Significant positive correlation was observed between 25(OH)D levels and aerobic performance (VO<sub>2</sub>max) and training status. Supplementation with supraphysiological dose of vitamin D (6000 IU/day) during 8-week of training in rowers with sufficient 25(OH)D levels significantly increased VO<sub>2</sub>max compared to placebo group <sup>25</sup>. However, no significant effect of vitamin D on athletic performance or association between 25(OH)D levels and an individual's VO<sub>2</sub>max were also noted <sup>40, 41</sup>.

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367 Several mechanisms have been reported that may be responsible for the protective and ergogenic effect of 25-hydroxycholecalciferol in skeletal muscle<sup>13</sup>. The proposed 368 369 mechanisms include a role of vitamin D receptors (VDR) that are expressed in skeletal 370 muscle and when bound to  $1,25(OH)_2D_3$ , exert genomic effects at target sites <sup>24</sup>. Another 371 mechanism includes a role of supplementation with vitamin D in stimulating oxygen uptake 372 in skeletal muscle. It has been hypothesized that positive effects of 25(OH)D on oxygen 373 uptake could be due to the fact that the cytochrome enzymes that activate vitamin D into 374 1,25-dihydroxycholecalciferol have heme-containing proteins that could potentially affect the binding affinity of oxygen to hemoglobin <sup>42</sup>. A significant effect of both exercise 375 376 training and vitamin D supplementation on increased force and power output of skeletal

muscle perhaps in response to an enhanced cross-bridge cycling and muscular contraction
has also been suggested <sup>22, 43, 44</sup>.

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380 In our study we concluded that 25(OH)D production after vitamin D diet has a significant 381 effect on selected biomarkers of skeletal muscle damage and post exercise proinflammatory 382 cytokine levels. Significant negative correlation was observed between 25(OH)D 383 concentration and TN level and 25(OH)D and MB concentration in response to a vitamin D 384 diet. Importantly, the negative correlation was observed between 25(OH)D concentration 385 and CK activity during the 24h recovery period and TNFa levels. These support the 386 findings that lower serum levels of biomarkers of skeletal muscle damage and vitamin D 387 status improvement, might, in turn, have significantly decreased individual recovery time in 388 marathon runners. Lower levels of serum vitamin D have been associated with increased muscle weakness, fatigue and injury incidents <sup>45</sup>. Therefore, the ability to reduce fatigue 389 390 and decrease the recovery time is important for athletes who train at high and moderate 391 intensity with both concentric and eccentric muscle contraction more frequently. It was also 392 observed that during recovery 1,25-hydroxyvitamin D increases the myogenic 393 differentiation and proliferation, down-regulates myostatin and improved the skeletal muscle regeneration in animal studies <sup>17</sup>. The findings that vitamin D supplementation 394 enhances the recovery process following intense exercise <sup>18</sup> and ultramarathon runs<sup>46</sup> were 395 396 also supported by human studies. Serum 25(OH)D concentrations correlated positively with 397 physical activity scores, and negatively with body mass index, lipid profile, fatigue scores (visual analog scale), and muscle fatigue biomarkers in healthy older adults <sup>47, 48</sup>. Higher 398 399 25(OH)D levels were accompanied by lower creatine kinase, troponin I, and lactic acid 400 dehydrogenase activity, the generally used biomarkers for earlier detection of muscle 401 injury, especially muscle soreness following training interventions <sup>34</sup>. In the study of 402 Nowak et al., self-reported fatigue has been linked to low levels of circulating 25403 hydroxyvitamin D (250HD), a biomarker of vitamin D status, however, vitamin D
404 treatment significantly improved fatigue in healthy persons with vitamin D deficiency <sup>47</sup>.
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406 Fatigue is a complex and nonspecific phenomenon with significant response to physical and 407 mental exertion or a feature of illnesses. There is no generally accepted set of criteria for fatigue, and the prevalence of fatigue varies widely depending on the assessment method <sup>49-</sup> 408 <sup>52</sup>. A previous study demonstrated that vitamin D supplementation attenuated the 409 410 inflammatory biomarkers immediately following intensive exercise with both eccentric and concentric muscle contractions <sup>19</sup>. Our results revealed lower post exercise TNF- $\alpha$  levels 411 412 and a tendency towards lower IL-6 concentrations in a specifically trained supplementation 413 group compared to the baseline levels. Regardless of the fact that long-term exercise 414 training might diminish 25(OH)D concentrations, we conclude that a dietary vitamin D 415 supplementation also has a beneficial effect on the function of the immune system by 416 suppressing exercise-induced proinflammatory cytokines in elite athletes. Still, a question 417 arises whether the recommended dosage of 1500-2000 IU/day vitamin D could maintain 418 adequate serum vitamin D concentrations in endurance trained athletes. The optimal levels 419 needed for athletic performance are controversial; lower than 1000 UI/day may not be 420 sufficient, especially for an older athletic population. It has been shown that dosages higher 421 than 2000 UI/day or 3000 UI/day have been sufficient to increase skeletal muscle function and reduce the risk of stress fractures <sup>23, 53, 54</sup>. The possible mechanisms responsible with a 422 423 detailed characteristic of skeletal muscle functions in response to different dosages of 424 vitamin D diet were not a major issue of the paper. These preliminary findings highlight the 425 requirement for further studies on the effects of different dosages of vitamin D 426 supplementation on skeletal muscle function and optimal performance in athletes.

428	In summary, our results show that a 3-week vitamin D supplementation had a beneficial
429	effect on skeletal muscle adaptation to running exercise with eccentric muscle contraction.
430	The improvement of muscle function and recovery observed in our study population might
431	have been induced by a decrease in biomarkers of muscle damage and injury associated
432	with higher serum 25(OH)D concentrations a vitamin D-rich diet.
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#### 450 **Declarations**

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#### 452 Ethics approval and consent to participate

- 453 The experiment was approved by the Ethics Committee of the Academy of Physical
- 454 Education in Katowice (Ethics Committee decision KBN 3.2016) and conformed to the
- 455 standards set by the Declaration of Helsinki.

456

457	Consent	for	publ	lication

458 All authors gave their consent for publication

459

- 460 Availability of data and material
- 461 Upon request from the first author
- 462
- 463 **Competing interests**
- 464 The authors declare no conflict of interest.

465

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- 469 Authors' contributions
- 470 Conceptualization, A.Ż. and Z.W.; Methodology, A.Z., E.S-K.; Validation, A.S..; Formal
- 471 Analysis, A.Z.; Investigation, O.Ł. A.Z, E. S-K; Data Curation, T.R., P.N. and B.K.;

- 472 Writing Original Draft Preparation, A.Z.; Writing Review & Editing, E.B., T.R., P.N.
- 473 and B.K.; Visualization, T.R., P.N. and B.K.; Supervision, T.R., P.N. and B.K.; Project
- 474 Administration, T.R., P.N. and B.K.
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Variables	EXP	CON
	<i>n</i> =12	<i>n</i> =12
Age (years)	$33.7\pm7.5$	$35.9\pm5.3$
Body mass (kg)	$74.7 \pm 10.6$	$75.3\pm8.6$
Body Height (cm)	$176.8\pm6.0$	$178.2 \pm 6.8$
BMI (kg/m <sup>2</sup> )	$23.8\pm2.2$	$23.7\pm2.1$
FAT (%)	$13.7 \pm 3.3$	$13.5 \pm 4.4$
SMM (kg)	$36.5 \pm 5.1$	$36.9\pm4.5$
TBW (L)	$47.2\pm6.4$	$47.5\pm5.4$
VO <sub>2</sub> max (mL/kg/min)	$54.5\pm9.4$	$54.5\pm9.4$
Peak power (Watt)	$321.5\pm77.9$	351.4 ± 68.3
HR max (b/min)	$181.0 \pm 11.0$	$186.0 \pm 9.0$

BMI- body mass index, FAT- percent of body fat, SMM – skeletal muscle mass, TBW – total body water,
 VO<sub>2</sub>max – maximal oxygen uptake, HR max – heart rate maximum.

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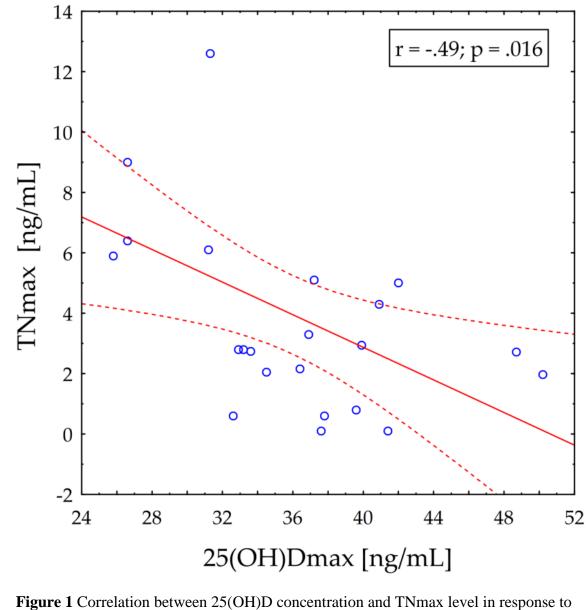
### Table 1 Subject characteristics (mean, SD)

Table 2 Mean energy supply with diet, mean daily fat, carbohydrate, protein and vitamin D
 intake in the supplemented group and placebo group (mean, SD).

Variables	EXP	CON
	( <i>n</i> =12)	( <i>n</i> =12)
Energy [kcal/kg/day]	$29.6\pm3.0$	$28.0\pm2.0$
Fat intake [%]	$31.7\pm9.6$	$30.8\pm8.3$
Carbohydrate intake [%]	$46.1\pm6.6$	$46.7\pm8.5$
Protein intake [%]	$22.8\pm5.4$	$22.4\pm3.3$
Vitamin D [µg/day]	$7.8\pm7.1$	$8.4 \pm 7.3$

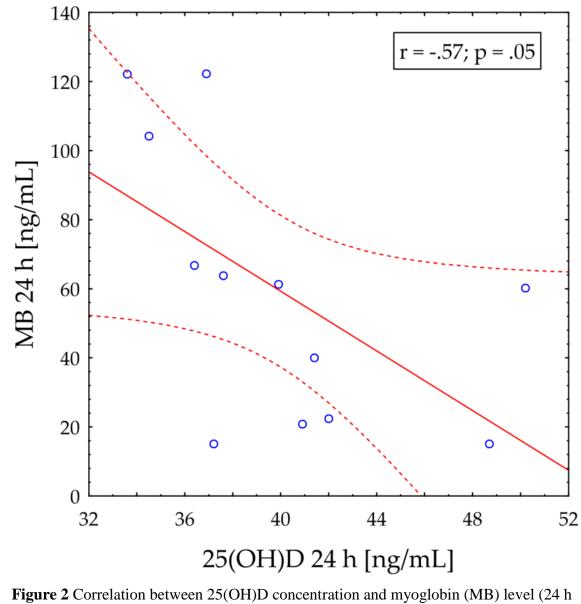
Variables	E	EXP		CON		Effect size
	Pre-Suppl	Post-Suppl	Pre-Placebo	Post-placebo	Post Suppl vs post Placebo	Cohen d
25(OH)rest [ng/ml]	$34.9\pm4.7$	40.3 ± 4.9 *	$33.9\pm4.8$	$31.8\pm4.2$	0.05	1.86 / Large
25(OH)max [ng/ml]	$36.5\pm3.3$	44.9 ± 4.9 ***	$34.7\pm8.1$	$39.2\pm7.6$	ns	0.89 / Moderate
25(OH)D1 h [ng/ml]	$40.0\pm8.8$	$45.5\pm4.7$	$33.3 \pm 3.4$	$38.5\pm9.7$	0.05	0.92 / Moderate
25(OH)D24h [ng/ml]	$36.2\pm6.2$	41.2 ± 5.0 *	$30.0\pm6.4$	$35.7\pm6.9$	0.05	0.91 / Moderate
TN rest [ng/ml]	$2.9\pm1.9$	$2.0 \pm 1.6$	$7.2 \pm 1.9$	$5.6 \pm 4.2$	0.05	1.13 / Moderate
TN max [ng/ml]	$5.1 \pm 1.7$	$2.7\pm1.6$	$8.9\pm 6.2$	$5.3 \pm 4.1$	0.001	0.84 / Moderate
TN 1 h [ng/ml]	$4.9\pm2.0$	$2.9 \pm 2.0 *$	$4.4\pm3.2$	$4.7\pm2.4$	ns	0.81 / Moderate
TN 24 h [ng/ml]	$6.3 \pm 3.7$	3.7 ± 1.2 *	$3.1 \pm 1.2$	$3.1 \pm 1.2$	ns	0.5 / Small
MB rest [ng/ml]	$44.7\pm23.1$	$40.6\pm17.6$	$44.4 \pm 11.8$	37.1 ± 21.8	ns	0.18 / Trivial
MB max [ng/ml]	$73.9\pm32.0$	$58.7\pm27.6$	$93.4\pm33.1$	$73.5\pm43.7$	ns	0.4 / Small
MB 1h [ng/ml]	$173.6\pm104.5$	$92.6\pm48.9$	$102.6\pm59.5$	$83.9\pm50.0$	ns	0.18 / Trivial
MB 24h [ng/ml]	$93.2\pm56.2$	59.5 ± 37.8 ***	$98.3\pm26.7$	$93.0\pm50.7$	ns	0.75 / Moderate
CK rest [U/l]	$151.0\pm59.5$	$166.4\pm95.5$	$234.2\pm88.9$	$248.4 \pm 179.0$	ns	0.57 / Small
CKmax [U/l]	$226.1\pm141.0$	$212.7 \pm 112.0$	$276.2 \pm 118.2$	$286.6\pm191.5$	ns	0.47 / Small
CK 1h [U/l]	$248.0\pm161.8$	$214.3 \pm 109.0$	$276.8 \pm 122.3$	$213.2 \pm 113.4$	ns	0.01 / Trivial
CK 24 h [U/l]	$361.3\pm228.9$	243.3 ± 91.5 *	$434.3 \pm 143.9$	$332.0 \pm 255.6$	0.05	0.46 / Small
LDH rest [U/l]	$337.1 \pm 73.5$	$333.1 \pm 80.5$	$339.4\pm47.8$	333.1 ± 60.1	ns	0 / Trivial
LDH max [U/l]	$400.5\pm108.0$	$395.9 \pm 68.6$	$401.4 \pm 63.8$	413.5 ± 79.6	ns	0.24 / Small
LDH 1h [U/l]	$361.4\pm87.8$	$354.2\pm69.4$	$355.0\pm44.9$	$368.6\pm72.2$	ns	0.2 / Small

LDH 24h [U/l]	$344.9\pm75.5$	$313.5\pm66.6$	$339.1\pm56.8$	321.1 ± 31.1	ns	0.15 / Trivial
TNFα rest [pg/ml]	$9.7 \pm 5.7$	$5.6 \pm 2.6$	$13.7\pm7.4$	$12.5 \pm 4.4$	ns	1.91 / Large
TNFα max [pg/ml]	$23.9 \pm 15.2$	10.5 ± 4.6 **	$22.9 \pm 13.7$	$22.7 \pm 17.4$	ns	0.96 / Moderate
TNFα 1h [pg/ml]	$21.9 \pm 16.8$	8.4 ± 3.7 **	$18.7 \pm 11.4$	$21.3 \pm 12.2$	ns	1.43 / Large
TNFα 24h [pg/ml]	$19.8 \pm 14.2$	$11.6 \pm 5.7$	$13.9\pm6.7$	$13.7\pm7.3$	ns	0.32 / Small
IL-6 rest [pg/ml]	$1.4 \pm 1.3$	$1.9 \pm 1.8$	$1.5 \pm 1.3$	$2.2 \pm 2.0$	ns	0.16 / Trivial
IL-6 max [pg/ml]	$2.0 \pm 1.9$	$1.7 \pm 1.0$	$2.7 \pm 1.5$	$2.5 \pm 2.3$	ns	0.45 / Small
IL-6 1h [pg/ml]	$2.7 \pm 2.3$	$2.3 \pm 1.3$	$3.1 \pm 2.0$	$3.0 \pm 1.9$	ns	0.43 / Small
IL-6 24h [pg/ml]	$1.8 \pm 1.2$	$1.0 \pm 0.9$	$2.0 \pm 1.2$	$2.4 \pm 1.6$	ns	1.08 / Moderate

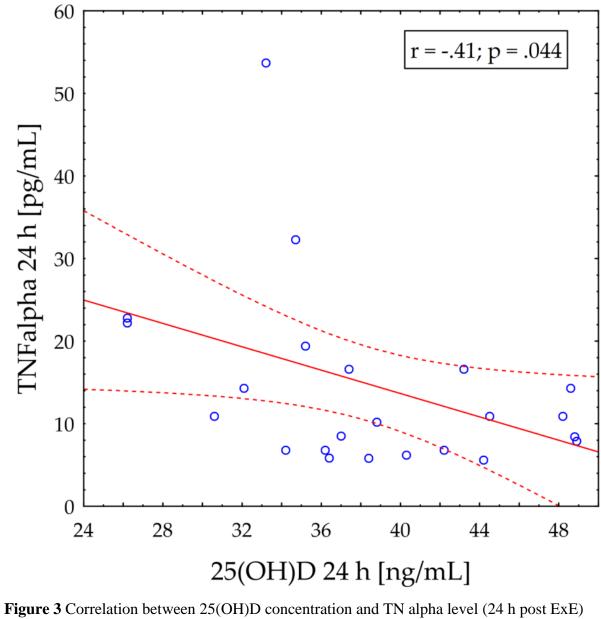




vitamin D supplementation.



tion between 25(OH)D concentration and myoglobin (MB) level (24 post ExE) in response to vitamin D supplementation.



676 in response to vitamin D supplementation.